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Susceptibility to anti-infective drugs of aerobic bacteria and fungi isolated from CASP: retrospective study in a pediatric tertiary care center

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ABSTRACT:

- Background: Perforated appendicitis is the most important cause of community acquired secondary peritonitis (CASP) in children. Knowledge of antimicrobial susceptibility of isolated pathogens is mandatory for a correct administration of empiric therapy.
- **Patients and Methods:** Retrospective analysis of pathogens isolated from peritoneal fluid in children with CAPS followed at the Istituto Giannina Gaslini (Genoa, Italy) from 2013 to 2015.
- **Results:** A total of 117 bacterial strains were detected: 52% Enterobacteriaceae, 14% P. aeruginosa, 19% Gram-positives, 13% Gram-negative anaerobes, and 2% Fungi. Resistance to ampicillin-sulbactam was detected in 33% of Enterobacteriaceae and 10% of enterococci. Moreover 15% of Enterobacteriaceae were resistant to gentamycin and 10% to piperacillin-tazobactam. No strain of P. aeruginosa was resistant to piperacillin-tazobactam. None of the Gram-negatives was resistant to cerbapenems and no Gram-positive strain was resistant to vancomycin. Concomitant resistance to ampicillin-sulbactam and gentamycin was detected in 13% of strains, while no strain was concomitantly resistant to piperacillin-tazobactam and gentamycin. These data were different from a previous 6-year observation in our centre.
- **Conclusions:** The present study demonstrates that continuous monitoring of aetiology and antimicrobial susceptibility is mandatory for a correct management of CASP in children.
- **Keywords:** CASP, Paediatric, Peritonitis, Drug-susceptibility, Piperacillin-tazobactam.

INTRODUCTION

Intra-abdominal infection (IAI) comprises a wide spectrum of pathological conditions, ranging from uncomplicated appendicitis to fecal peritonitis (intra-abdominal abscesses or diffuse peritonitis) (1). IAI may be classified according to whether the infection is community acquired (approximately 80%) or health care associated, the latter being the most common complication of elective or emergency intra-abdominal surgery. In children the most important causes of secondary peritonitis are per-

forated appendicitis and intra-abdominal abscess arising from acute appendicitis¹. The etiology of this disease is predominantly related to the organisms of the gut flora such as *Enterobacteriaceae*, enteric Gram-positive *cocci*, and obligate anaerobes^{2,3}. The exact role of *Pseudomonas aeruginosa* in this condition remains poorly clarified, even if it represents a non negligible proportions of pathogens isolated in pediatric secondary peritonitis⁴. Studies of antibiotic susceptibility of pathogens isolated in Community Acquired Secondary Peritonitis (CASP) in children are limited^{1,5,6}. These issues are critical in an

attempt to choose empirical, initial antibiotic treatment of secondary peritonitis. The most frequently suggested treatments are represented by monotherapy with a carbapenem or the combination of a penicillin with a beta-lactamase inhibitors like piperacillin-tazobactam or ticarcillin-clavulanate, or amoxicillin-clavulanate, ertapenem or piperacillin-tazobactam in absence of extended-spectrum-beta-lactamase (ESBL) producing strains. Meropenem or imipenem-cilastatin is recommended in case of suspicion of infections due to resistant strains^{7,8}. In any case the choice of therapy, both for children and adults, should also be driven by severity of clinical condition, and/or local epidemiological data^{8,9}.

In 2013, a standard operative procedure for the initial empirical treatment of CASP was established in our hospital¹⁰. The choice of antibiotics was based on antimicrobial susceptibility of pathogens isolated from CASP in the 6 previous years, and the combination of ampicillinsulbactam and gentamycin was suggested. At the same time, a prospective survey on the number of re-admission to the surgical ward of patients previously undergoing surgery for CASP was implemented and showed a 13.5% of re-interventions compared with the 5.6% of the previous three years. Surgical procedures were reviewed as well as microbiology and antimicrobial susceptibility of peritoneal isolated after 2013 in order to update the recommendations. The aim of the present study is to report changes in antimicrobial susceptibility of aerobic bacteria isolated from peritoneal cultures in children with CASP in the period January 2013 – December 2015.

PATIENTS AND METHODS

Patients

Istituto Giannina Gaslini (IGG), Genoa - Italy is a Tertiary Care Pediatric Hospital in Northern Italy serving as local Pediatric Hospital for the Genoa area, and as tertiary care referring hospital for Italy and for many foreign countries. Clinical records of patients diagnosed with community acquired secondary peritonitis from January 2013 to December 2015 were retrospectively reviewed in order to collect data on pathogens isolated from the peritoneal fluid and

their susceptibility to anti-infective drugs. The diagnosis of CASP was based on 1) attending surgeon judgment based on the presence of transmural necrosis of the appendix, a defect in the wall of the appendix, an abscess cavity, or diffuse purulent fluid at the time of operation and 2) the absence of any risk factor (e.g. immunocompromission, inflammatory bowel disease, peritoneal dialysis, recent abdominal surgery), as reported on patient's clinical record.

Methods

Cultures were performed on peritoneal fluid sampled immediately after peritoneal incision. The peritoneal fluid was inoculated into both aerobic and anaerobic bottles (BactAlert Biomerieux, France) in the operating theatre, transferred to the laboratory within 1 hour after collection and incubated at 35°C under continued automated monitoring for seven days (BactAlert Detection System). Positive samples were Gram-stained and subcultured on agar plates according to laboratory standard methods¹¹. Bacteria identification and antibiotic susceptibility tests (AST) were performed by automatic system (BD Diagnostics Systems, Franklin Lakes, NJ, USA) according to the manufacturer's recommendations. The ASTs were expressed as minimal inhibitory concentrations (MICs) and interpreted using EUCAST breakpoint tables¹². Amoxicillin-clavulanate was substituted with ampicillin-sulbactam since only this drug was available for intravenous therapy in our Institute. AST for anaerobes was not evaluated. Antifungal susceptibility was evaluated by means of the YeastOne Sensititre (Trek Diagnostic System Ltd. West Sussex, UK).

RESULTS

During the study period, 117 strains were retrieved from cultures of peritoneal fluid in 76 patients with CASP. The patients median age was 7.8 years (range: 3-12 years), 40 were male (52%), while 36 (48%) were female. The majority of the patients were Italians (60; 79%) while 16 were foreign-born children (16; 21%). Cultures resulted polymicrobial in 42 (55%) cases

Table 1. Microorganisms Recovered From Peritoneal Fluid in children with community-acquired secondary peritonitis.

| | Microorganisms | Number of strains (%) | |
|-----------|--|-----------------------|--|
| Aerobes | Gram-negatives | 77 (66) | |
| | E. coli | 53 (45) | |
| | P. aeruginosa | 16 (14) | |
| | Klebsiella-Enterobacter-Serratia group | 7 (6) | |
| | P. mirabilis | 1 (<1) | |
| | Gram-positives | 23 (19) | |
| | Streptococci (S.constellatus, S.anginosus) | 13 (11) | |
| | E.faecalis | 2 (2) | |
| | E.faecium | 5 (4) | |
| | Enterococcus sp | 3 (2) | |
| Anaerobes | Bacteroides spp | 15 (13) | |
| Fungi | | 2(2) | |
| Total | | 117 (100) | |

and monomicrobial in 34 (45%). Table 1 reports the proportions of different isolated pathogens. The most frequently isolated aerobe was *Escherichia coli* (45%), while *Pseudomonas aeruginosa* represented 14% of the identified strains. *Enterococci* represented 8% of the identified strains, Gram-negative anaerobes accounted for 13% of the identified strains, while 2% were *Candida albicans*.

Table 2 reports on antibiotic susceptibility of pathogens isolated from peritoneal cultures. Resistance to ampicillin-sulbactam was detected in 33% of tested strains, and considering the absence of activity of this drug against *Pseudomonas aeruginosa*, the total of resistant strains to ampicillin-sulbactam was 44 out of 100 aerobe strains identified. Moreover, in a further 27% (n=14) of tested strains the MIC was at the breakpoint. Resistance to gentamycin was detected in 9% of Gramnegative aerobes, but in another 7% (n=6) of the cases the MIC was at the breakpoint. As for piperacillin-tazobactam 91% of strain tested resulted susceptible. It is noteworthy that the highest proportion of resistant strains (29%) was found in the Klebsiella-Enterobacter-Serratia group. None of the tested strains was resistant to meropenem, with all MIC < 2 mg/L. Similar results were observed for ertapenem that was effective against all the *Enterobacteriaceae*, in all cases with MIC < 0.5 mg/L. Finally, none of the enterococci was gentamycin or vancomycin-resistant, but 1 (10% of enterococci) strain was resistant to ampicillin.

Concomitant resistance to ampicillin-sulbactam and gentamycin was detected in 14% of *Enterobacteriaceae* (n=7), while no strain was concomitantly resistant to piperacillin-tazobactam and gentamycin. As for fungal pathogens, both strains of *Candida albicans* isolated from peritoneal fluid were resistant to fluconazole, but susceptible to amphotericin B and micafungin. No data for antibiotic susceptibility of anaerobes were available.

DISCUSSION

Perforated appendicitis and intra-abdominal abscess arising from acute appendicitis represent the most important cause of CASP in children. In our series, Escherichia coli and Pseudomonas aeruginosa followed by Bacteroides were the most frequently isolated organisms involved in CASP, even if these proportions were different from those observed in previous pediatric studies, as summarized in Table 36,10,13-16. These differences might be attributable, at least in part, to differences among the populations evaluated in in previous studies, since our patients were affected only by appendicitisinduced peritonitis, while in some other reports were considered all patients with intra-abdominal infections requiring surgery. Knowledge of the bacterial susceptibility to antibiotics is critical in an attempt to provide the best antibiotic treatment since the beginning. The more recent recommendations have suggested monotherapy with ampicillin-sulbactam or ertapenem or piperacillintazobactam for primary empirical treatment in absence of risk factors for infections by resistant pathogens⁷⁻⁹.

Table 2. Distribution of susceptibility to different antibiotics in children with advanced peritonitis

| Microorganism (n=number of tested strains) | Minimal Inhibitory Concentration (mg/L) and interpretation (n=number of strains)* | E. coli n=53** | P. aerugi- nosa n=16 | Klebsiella- Enterobacter- Serratia group n=7** | Proteus spp. n=1** | E. faecalis n=2** | E. faecium n=5** | Enterococcus spp. n=3 (**) | Streptococci n=13** |
|--|---|--------------------|-------------------------|---|-----------------------|----------------------|---------------------|-------------------------------|------------------------|
| Ampicillin-sulbactam (n= 84) | <pre><8 (S) n=56 (67) >8 (R) n=28 (33)</pre> | 30 (57) 23 (43) | | 3 (43) 4 (57) | 1 (100) | 2 (100) | 4 (80) 1 (20) | 3 (100) | 13 (100) |
| Gentamicin (n=87) | <pre><4 (S) n=79 (91) >4 (R) n=8 (9)</pre> | 45 (85) 8 (15) | 16 (100) | 7 (100) | 1 (100) | 2 (100) | 5 (100) | 3 (100) | |
| Piperacillin-tazobactam (n=77) | <pre><16 (S) n=70 (91) >16 (R) n=7 (9)</pre> | 48 (90) 5 (10) | 16 (100) | 5 (71) 2 (29) | 1 (100) | | | | |
| Meropenem (n=77) | <8 (S) n=77 (100) >8 (R) n=0 | 53 (100) | 16 (100) | 7 (100) | 1 (100) | | 1 1 | 1 1 | 11 |
| Ertapenem (n=54) | ≤ 1 (S) n=54 (100) >1 (R) n=0 | 53 (100) | | | 1 (100) | | | | |
| Vancomycin (n=10) | <pre><4 S n=10 (100) >4 (R) n=0</pre> | | | | 1 1 | 2 (100) | 5 (100) | 3 (100) | |

*Numbers in parenthesis are proportions of the total number of tested strains.
** Numbers in parenthesis are proportions of the total number of isolated strains

Table 3. Proportions of bacteria isolated from CASP in children.

| Study/Year of publication (Reference) | Present study | Castagnola E et al ¹⁰ | Guillet- Caruba et al ¹⁵ | Dumont et al ¹⁶ | Lin et al ⁶ | Bennion* et al ¹⁴ | Brook et al ¹³ |
|---|------------------|-------------------------------------|---|-------------------------------|---------------------------|---------------------------------|------------------------------|
| Number of patients | 76 | 106 | 86 | 70 | 100 | 30 | 113 |
| Number of strains | 117 | 114 | 136 | 123 | 445 | 305 | 325 |
| Proportions of isolated ba | ecteria | | | | | | |
| E. coli | 45 | 74 | 71 | 51 | 31 | 82 | 13 |
| Other Enterobactericeae | 6 | 8 | 7 | 4 | 11 | 10 | 4.7 |
| P. aeruginosa | 14 | 13 | 19 | 6 | 6 | 23 | 2 |
| Bacteroides spp | 13 | _ | 20 | 22 | 9 | 73 | 23 |
| Streptococci | 11 | _ | 34 | 9 | 18 | 33 | 12 |
| Enterococci | 8 | 6 | 8 | 1 | 3 | 33 | 2.6 |

^{*}all polymicrobial infection.

In our previous study we documented that these recommendations could be not suitable for children or at least in all pediatric centers¹⁰. In the present study, we also showed that continuous evaluation of epidemiology and antimicrobial susceptibility is mandatory, since the combination ampicillin-sulbactam + gentamycin that in the period 2005-2011 had a potential 99% effectiveness, 2 years later was effective only in 86% of isolated strains (-13%)¹⁰. One of the most important statements of the guidelines for the treatment of peritoneal infections is that "if resistance to a given antibiotic is present in 10-20% or more of isolates of a common intra-abdominal pathogen in the community, use of that agent should be avoided⁷⁻⁹". According to this advice, we decided to change from the combination with ampicillinsulbactam and gentamycin to piperacillin-tazobactam and gentamycin that allowed a potential effectiveness of 99-100%¹⁷. This decision was also made taking into account the high proportion of *Pseudomonas aerugino*sa we observed in the present study and in our previous survey¹⁰.

CONCLUSIONS

The high proportion of *Pseudomonas aeruginosa* isolated in peritonitis is a peculiarity of pediatric CASP and must not be neglected^{7,18-21}. The use of piperacillin-tazobactam and gentamycin is correct also taking in consideration the antibiotic susceptibility of other pathogens like *enterococci* (10% resistance to ampicillin, 0% resistance to gentamycin) and *streptococci* (100% sensitivity to ampicillin). Even if not evaluated, piperacillin is effective also against anaerobes giving another reason for choosing it as a CASP first-line empiric treatment.

CONFLICTS OF INTEREST:

The authors declare no conflict of interest.

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REFERENCES

- 1. Menichetti F, Sganga G. Definition and classification of intraabdominal infections. J Chemother 2009; 21: 3-4.
- Kaplan S. Antibiotic usage in appendicitis in children. Pediatr Infect Dis J 1998; 17: 1047-1048.
- 3. Chen CY, Chen YC, Pu HN, Tsai CH, Chen WT, Lin CH. Bacteriology of acute appendicitis and its implication for the use of prophylactic antibiotics. Surg Infect (Larchmt) 2012; 13: 383-390.
- Obinwa O, Casidy M, Flynn J. The microbiology of bacterial peritonitis due to appendicitis in children. Ir J Med Sci 2014; 183: 585-591.
- 5. Solomkin JS, Mazuski JE, Bradley JS, Rodvold KA, Goldstein EJ, Baron EJ, O'Neill PJ, Chow AW, Dellinger EP, Eachempati SR, Gorbach S, Hilfiker M, May AK, Nathens AB, Sawyer RG, Bartlett JG. Diagnosis and management of complicated intra-abdominal infection in adults and children: guidelines by the Surgical Infection Society and the Infectious Diseases Society of America. Clin Infect Dis 2010; 50: 133-164. Erratum in: Clin Infect Dis 2010; 50: 1695. Dosage error in article text.
- Eckmann C, Dryden M, Montravers P, Kozlov R, Sganga G. Antimicrobial treatment of "complicated" intra-abdominal infections and the new IDSA guidelines? A commentary and an alternative European approach according to clinical definitions. Eur J Med Res 2011; 16: 115-126.
- Decousser JW, Lamy B, Pina P, Allouch PY; Collège de Bactériologie Virologie Hygiène Study Group (ColBVH). Trends in antibiotic susceptibility of bloodstream pathogens in hospitalized patients in France, 1996 to 2007. Diagn Microbiol Infect Dis 2010; 66: 292-300.
- Lin WJ, Lo WT, Chu CC, Chu ML, Wang CC. Bacteriology and antibiotic susceptibility of community-acquired intraabdominal infection in children. J Microbiol Immunol Infect 2006: 39: 249-254.
- Sartelli M, Viale P, Koike K, Pea F, Tumietto F, van Goor H, Guercioni G, Nespoli A, Tranà C, Catena F, Ansaloni L, Leppaniemi A, Biffl W, Moore FA, Poggetti R, Pinna AD, Moore EE. WSES consensus conference: guidelines for first-line management of intra-abdominal infections. World J Emerg Surg 2011; 6: 2.
- 10. Castagnola E, Bandettini R, Ginocchio F, Perotti M, Masa DL, Ciucci A, Loy A, Caviglia I, Haupt R, Guida E, Pini Prato A, Mattioli G, Buffa P. Susceptibility to antibiotics of aerobic bacteria isolated from community acquired secondary peritonitis in children: therapeutic guidelines might not always fit with and everyday experience. J Chemother 2013; 25: 213-216.

- Murray PR. Processing and interpretation of sterile body fluids. Manual of Clinical Microbiology, Vol. 1. 6th ed. Washington, DC: ASM Press; 1995.
- European Committee on Antimicrobial Susceptibility Testing (EUCAST), http://www.eucast.org; 2016 (accessed 10.20.2016).
- 13. Brook I. Bacterial studies of peritoneal cavity and postoperative surgical wound drainage following perforated appendix in children. Ann Surg 1980; 192: 208-212.
- Bennion RS, Baron EJ, Thompson JE Jr, Downes J, Summanen P, Talan DA, Finegold SM. The bacteriology of gangrenous and perforated appendicitis--revisited. Ann Surg 1990; 211: 165-171.
- Guillet-Caruba C, Cheikhelard A, Guillet M, Bille E, Descamps P, Yin L, Khen-Dunlop N, Zahar JR, Sarnacki S, Révillon Y, Berche P, Ferroni A. Bacteriologic epidemiology and empirical treatment of pediatric complicated appendicitis. Diagn Microbiol Infect Dis 2011; 69: 376-381.
- Dumont R, Cinotti R, Lejus C, Caillon J, Boutoille D, Roquilly A, Podevin G, Gras-Le Guen C, Asehnoune K. The microbiology of community-acquired peritonitis in children. Pediatr Infect Dis J 2011; 30: 131-135.

- Lovering AM, Reeves DS. Aminoglycosides and aminocyclitols. In: Finch RG, Greenwood D, Norrby SR, Whitley RJ, editors. Antibiotic and chemotherapy. London: Sauders Elvisier; 2010, pp. 145-169.
- Yellin AE, Heseltine PN, Berne TV, Appleman MD, Gill MA, Riggio CE, Chenella FC. The role of Pseudomonas species in patients treated with ampicillin and Sulbactam for gangrenous and perforated appendicitis. Surg Gynecol Obstet 1985; 161: 303-307.
- Aronoff SC, Olson MM, Gauderer MW, Jacobs MR, Blumer JL, Izant RJ Jr. Pseudomonas aeruginosa as a primary pathogen in children with bacterial peritonitis. J Pediatr Surg 1987; 22: 861-864.
- Bradley JS, Behrendt CE, Arrieta AC, Harrison CJ, Loeffler AM, Iaconis JP, Wald ER. Convalescent phase outpatient parenteral antiinfective therapy for children with complicated appendicitis. Pediatr Infect Dis J 2001; 20: 19-24.
- Maltezou HC, Nikolaidis P, Lebesii E, Dimitriou L, Androulakakis E, Kafetzis DA. Piperacillin/Tazobactam versus cefotaxime plus metronidazole for treatment of children with intra-abdominal infections requiring surgery. Eur J Clin Microbiol Infect Dis 2001; 20: 643-646.